

Research advances in understanding the role and mechanism of pyroptosis in myocardial ischemia-reperfusion injury

Ziyue Li¹, Bailong Hu², Xiaohua Zou²

¹College of Anesthesiology, Guizhou Medical University, Guizhou 550004, China. ²Department of Anesthesiology, the Affiliated Hospital of Guizhou Medical University, Guizhou 550004, China.

Corresponding authors: Bailong Hu and Xiaohua Zou.

Acknowledgement: This work was supported by the National Natural Science Foundation of China (No. 82160951, 82160224), the project of Guiyang Science and Technology Plan (zhukehe[2024]-2-27), the Cultivate project 2021 for National Natural Science Foundation of China, the Affiliated Hospital of Guizhou Medical University (gyfynsfc-2021-35, gyfynsfc-2021-49).

Declaration of conflict of interest: None.

Received December 5, 2023; Accepted February 4, 2024; Published June 30, 2024

Highlights

Currently, ischemic heart disease ranks as the most prevalent form of primary heart disease. The risk of myocardial ischemia-reperfusion injury, along with its associated mortality, is notably rising among perioperative patients. Recognizing the underlying mechanisms of myocardial ischemia-reperfusion injury and identifying suitable treatments are crucial. Inhibitors targeting the key molecules involved in pyroptosis hold promise as potential therapeutic options for managing myocardial ischemia-reperfusion injury.

Abstract

Myocardial ischemia-reperfusion injury (MIRI) emerges when the restoration of blood flow fails to recover myocardial function following transient ischemia, marking a significant pathological challenge that adversely affects revascularization outcomes and patient mortality. This condition often occurs post-cardiac procedures, including cardiopulmonary bypass, angioplasty, primary percutaneous coronary intervention, and thrombolytic therapy. Over the last decade, researches have been pivotal in deciphering the pathophysiological underpinnings of MIRI, aiming to identify viable targets and therapeutics for mitigation. Among these, pyroptosis, a form of inflammatory, programmed cell death, has been recognized for its integral role in MIRI, interacting with various other mechanisms such as oxidative stress, calcium dysregulation, autophagy, ferroptosis, and apoptosis. This review delves into the mechanisms by which pyroptosis influences MIRI, discusses its impact on both cardiomyocytes and non-cardiomyocytes in MIRI, and highlights recent advancements in the development of inhibitors targeting key molecules involved in pyroptosis such as Nod-like receptor protein 3 inhibitors, Caspase-1 inhibitors, and traditional Chinese medicines.

Keywords: Myocardial ischemia-reperfusion injury, pyroptosis, Nod-like receptor protein 3

Introduction

Acute myocardial infarction (AMI) is characterized by acute myocardial necrosis resulting from various pathogenic factors, including coronary stenosis, coronary plaque rupture, and atherosclerosis. Due to its rapid progression, AMI can lead to a high incidence of severe

complications and morbidity if not addressed promptly, making it a leading cause of death among patients with coronary artery disease globally [1-3]. Coronary artery disease ranks as the third leading cause of death worldwide, contributing to approximately 17.8 million fatalities each year [4]. Early coronary artery revascularization is crucial in treating AMI and

Address correspondence to: Bailong Hu, Department of Anesthesiology, the Affiliated Hospital of Guizhou Medical University, NO. 28 Guiyi Street, Guiyang 550004, Guizhou, China. Phone number: +86-15185184309; E-mail: hucailong@gmc.edu.cn. Xiaohua Zou, Department of Anesthesiology, the Affiliated Hospital of Guizhou Medical University, NO. 28 Guiyi Street, Guiyang 550004, Guizhou, China. Tel: +86-13809416036; Fax: +86-851-86771013; E-mail: zouxiaohuazh@gmc.edu.cn.

minimizing infarct size. The main treatment strategies include percutaneous coronary intervention (PCI) and coronary artery bypass Grafting for managing ST-segment elevation AMI [5-7].

Nonetheless, a pathological condition known as myocardial ischemia-reperfusion Injury (MIRI) can occur, where, paradoxically, myocardial function fails to improve or even worsens after reperfusion, further aggravating the injury [8, 9]. MIRI often manifests immediately to within two hours post-PCI in myocardial infarction patients, with a marked decrease in postoperative diastolic function compared to pre-operative levels [10]. Factors such as the short duration between AMI onset and infarct-related artery revascularization, infarction in the inferior wall, reduced forward blood flow in the infarct-related artery (less than TIMI grade 1), multivessel lesions, and renal insufficiency heighten the risk of MIRI following primary PCI.

Conversely, pre-infarction angina might serve as a cardioprotective factor, mitigating the severity of MIRI [11]. MIRI is characterized by high morbidity and mortality and can lead to numerous complications, including myocardial stunning, the no-reflow phenomenon, and reperfusion arrhythmias [12-14]. MIRI significantly influences the efficacy of revascularization and the mortality among these patients. The currently acknowledged pathogenic mechanisms of MIRI encompass several theories and processes, including the leukocyte theory, calcium overload, elevated reactive oxygen species (ROS), endoplasmic reticulum stress, autophagy, and apoptosis [15]. However, the complete understanding of MIRI's mechanisms remains elusive, attributed to the myriad of contributing factors and the complexity of its underlying mechanisms.

Pyroptosis, a proinflammatory type of programmed cell death, has gained substantial interest in recent times. This process is triggered by inflammasomes and members of the cysteinyl aspartate-specific proteinase (caspase) family. It is marked by the rupture of the cell membrane and the consequent release of inflammatory factors, which intensify the inflammatory cascade. Pyroptosis accelerates many cardiovascular diseases, such as atherosclerosis, heart failure, myocardial infarction, and other disease processes [16, 17]. Therefore, elucidating the mechanism of pyroptosis may help alleviate MIRI by targeting this pathway.

Overview of pyroptosis

History of pyroptosis

Friedlander AM first documented that treating mouse macrophages with anthrax lethal toxin resulted in cell death accompanied by the release of cellular contents [18]. Following this discovery, Zychlinsky et al. observed that *Shigella flexneri* induced death in mouse macrophages through a mechanism akin to apoptosis [19]. Both studies highlighted a novel mechanism of cell death, distinct from traditional apoptosis, characterized by the secretion of cellular contents.

In 2001, D'Souza et al. introduced the term "pyroptosis", deriving from Greek roots to describe this phenomenon. Pyroptosis involves the rupture of the cell membrane and the release of inflammatory factors, intensifying the surrounding inflammatory response. This process visually resembles burning, hence the name "pyroptosis" [20]. As researches evolved, pyroptosis came to be defined as a form of proinflammatory cell death mediated by cysteine-aspartate-specific proteinase-1 (caspase-1), marking a significant advancement in our understanding of cell death mechanisms [21].

Following initial discoveries, subsequent researches identified that gasdermin protein D (GSDMD) serves as the ultimate substrate for caspase-1 and caspase-11/4/5, establishing pyroptosis as a form of programmed cell death predominantly triggered by the activation of GSDMD by the caspase family [22]. GSDMD acts as an executor, breaching the cell membrane to facilitate the release of inflammatory factors [16, 23]. In 2018, the Nomenclature Committee for Cell Death formally recognized pyroptosis as a type of regulatory programmed cell death. This process is characterized by the involvement of gasdermin protein family members, which perforate the cell membrane, leading to cell swelling and eventual rupture. This event is typically accompanied by caspase family activation and the consequent leakage of inflammatory factors, further elucidating the intricate mechanisms underlying pyroptosis [24].

Key molecules involved in apoptosis

Pyroptosis is orchestrated through the formation of inflammasomes, which trigger the cleavage and perforation of GSDMD and result in the proinflammatory cell death characterized by the release of interleukin-1 β (IL-1 β) and interleukin-18 (IL-18). The principal

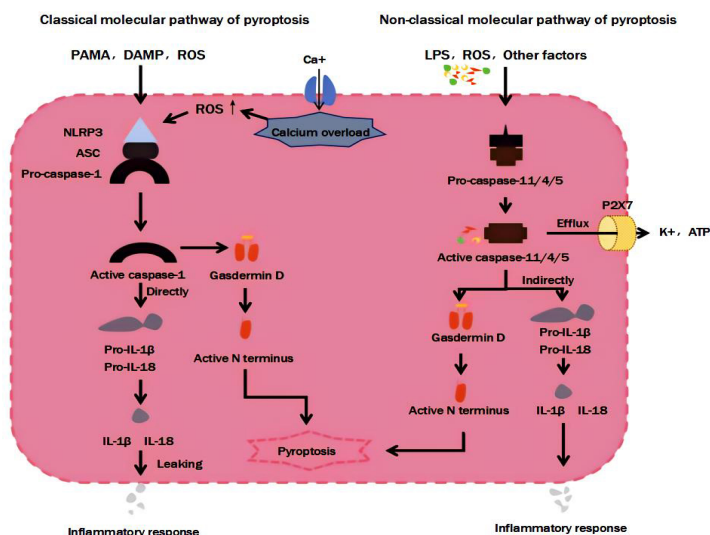


Figure 1. The process of pyroptosis. PAMP, pathogen-associated molecular pattern; DAMP, damage-associated molecular pattern; ROS, reactive oxygen species; LPS, lipopolysaccharide; ASC, apoptosis-associated speck-like protein; IL, interleukin; ATP, adenosine triphosphate.

proteins and molecules implicated in pyroptosis encompass the Nod-like receptor protein 3 (NLRP3) inflammasome, caspase-1, and GSDMD [25, 26].

Inflammasomes are multi-molecular complexes that are activated by various physical and chemical stimulus, such as pathogen-associated molecular patterns, ROS, and damage-associated molecular patterns [27, 28]. Activation occurs when such stimulus engages pattern recognition receptor on the cell surface, initiating a cascade of cell signaling events. This process leads to the activation of the inflammasome, which then orchestrates the assembly and activation of downstream signaling pathways, culminating in the release of inflammatory cytokines [29]. Inflammasomes encompass a variety of components, including CARD-domain containing 4, absent in melanoma 2, the Nod-like receptor (NLR) family pyrin domain containing NLRP1, NLRP3, and pyrin, among which NLRP3 is the most common.

The NLRP3 inflammasome is composed of three key elements: (1) the NLRP3 sensor molecule, (2) an apoptosis-associated speck-like protein (ASC) that includes a caspase-recruitment domain, and (3) the precursor form of cysteine aspartate-specific proteinase-1 (pro-caspase-1) [30]. NLRP3 facilitates the conversion of pro-caspase-1 into its active form, caspase-1, which plays a pivotal role in generating IL-18 and IL-1 β and in cleaving the GSDMD protein [31, 32]. GSDMD, a member of the gasdermin protein family, is expressed

across various cells and tissues in the cytoplasm. In its inactive state, GSDMD's N-terminus is bound to its C-terminal domain. Upon cleavage by caspase-1, the N-terminal fragment of GSDMD (GSDMD-N) separates and attaches to phosphoinositides on the cell membrane, subsequently perforating the membrane [33].

Classical molecular pathway of pyroptosis

The classical pathway of pyroptosis is orchestrated by Caspase-1 (See **Figure 1**). Upon exposure of pattern recognition receptors to pathogen-associated or damage-associated molecular patterns, NLRP3 becomes activated, facilitating the transformation of pro-caspase-1 into its active form, caspase-1. As previously discussed, GSDMD is subsequently cleaved by activated caspase-1 into two fragments: GSDMD-N and GSDMD-C. GSDMD-N forms pores ranging from 10–15 nm within the cell membrane's lipid layer, enhancing cell permeability. This increase in permeability leads to the leakage of intracellular inflammatory substances, amplifying the inflammatory response. Concurrently, the influx of extracellular fluid into the cells results in cell swelling, rupture and eventual lysis [34, 35]. Moreover, caspase-1 also cleaves pro-IL-1 β and pro-IL-18, activating them into IL-1 β and IL-18, respectively. These cytokines then escape through the pores formed by GSDMD-N, further propagating the inflammatory response in the surrounding tissue [36-38].

Non-classical molecular pathway of pyroptosis

The non-classical pathway of pyroptosis represents a divergence from the traditional caspase-1-mediated mechanism, relying instead on the activation of caspase-11/4/5 (See **Figure 1**). This pathway is triggered by bacterial lipopolysaccharide (LPS), ROS, and a variety of physical, chemical, and biological stimulus acting on caspase-11/4/5 [39]. Similar to caspase-1, activated caspase-11/4/5 can cleave GSDMD and facilitate the release of IL-1 and IL-18, contributing to the process of pyroptosis [40]. Additionally, caspase-11/4/5 plays a role in modulating the efflux of potassium (K⁺) and adenosine triphosphate via the P2X7 membrane channel, thereby

mediating pyroptosis in target cells [41]. It is important to note that while caspase-11/4/5 does not directly target pro-IL-1 β and pro-IL-18 for activation, it can indirectly influence and promote the release of these cytokines, further amplifying the inflammatory response associated with pyroptosis [42].

Other pathways of pyroptosis

Proteins within the gasdermin family exhibit highly conserved structural features, including C- and N-terminal domains, with the N-terminal domain acting as the functional executor responsible for inducing pyroptosis in target cells [43]. Traditionally, caspase-3/8 were not considered to impact GSDMD or to induce pyroptosis. However, current studies have revealed that certain molecular agents, such as chemotherapeutic drugs, can “trick” caspase-3 into cleaving Gasdermin E (GSDME), allowing the GSDME-N to perforate the cell membrane and effectively trigger pyroptosis in the target cell [33, 34].

Additionally, caspase-8, when influenced by factors like tumor necrosis factor-alpha, can cleave Gasdermin C (GSDMC) into GSDMC-N, which then perforates the cell membrane and mediates pyroptosis [44]. Liu et al. have also demonstrated that chimeric antigen receptor T cells can activate caspase-3 within target cells, leading to pyroptosis through the caspase-3/GSDME pathway [45]. Furthermore, the activation of the Gasdermin B (GSDMB) pathway has been shown to induce pyroptosis [46]. These novel mechanisms significantly broaden the understanding of pyroptosis, highlighting its complexity and the diverse roles of caspases and gasdermin proteins in cell death.

Role of pyroptosis in MIRI

Pyroptosis occurs in both cardiomyocytes and non-cardiomyocytes, significantly contributing to the progression of MIRI. During MIRI, the pyroptosis of various cell types, such as cardiomyocytes, fibroblasts, vascular endothelial cells, and macrophages, is triggered by mechanisms such as increased ROS, calcium overload, and an inflammatory response. This process intensifies the development of MIRI through multiple regulatory mechanisms, including mitochondrial dysfunction, altered energy metabolism, and oxidative stress. Pyroptosis interacts with MIRI and affect each other.

Xu et al. demonstrated that, in comparison to normal rats, the MIRI model group exhibited

a higher rate of myocardial cell pyroptosis, an enlarged area of myocardial infarction, and an intensified inflammatory response. However, the use of NLRP3 inhibitors or the genetic knockdown of NLRP3 was shown to mitigate myocardial pyroptosis, reduce infarct size, and lessen the inflammatory response [47]. Similarly, Sandanger et al. found that, during myocardial MIRI, the combined effects of ROS and potassium (K⁺) efflux could activate the NLRP3 inflammasome in cardiac fibroblasts, leading to the secretion of IL-1 β and the subsequent pyroptosis of these cells [48].

Furthermore, Zhang et al. reported that the surge of ROS during MIRI stimulates the NLRP3 inflammasome in myocardial microvascular endothelial cells, inducing pyroptosis and thereby aggravating inflammation and myocardial damage in the affected area [49]. Moreover, Dai et al. demonstrated that microRNA-2a, carried by exosomes derived from M2 macrophages (M2D-exos), can mitigate MIRI by targeting the TXNIP and TLR4/NF- κ B/NLRP3 signaling pathways [50]. In a related vein, Wang et al. observed that hypoxia/reoxygenation treatment prompts macrophages to adopt the M1 phenotype, which in turn mediates cardiomyocyte pyroptosis through miR-1a carried by exosomes [51]. Further research by Wang et al. uncovered that an increase in hypoxia-inducible factor-1 alpha can diminish MIRI by lowering mitochondrial ROS production and minimizing mitochondrial damage [52]. Additionally, Tian et al. found that preconditioning with isoflurane, especially when combined with captopril treatment over three days (as opposed to just one hour), significantly reduces MIRI by lessening oxidative stress and inflammation [53].

These findings collectively underscore the critical role of pyroptosis in both myocardial and non-myocardial cells, such as fibroblasts, vascular endothelial cells, and macrophages, is inextricably linked to MIRI. Thus, targeting pyroptosis across various cellular components emerges as a pivotal approach in the prevention and treatment of MIRI.

Relationship between pyroptosis and other MIRI-related mechanisms

Pyroptosis and oxidative stress

During MIRI, a significant increase in ROS occurs, especially in the early phases of reperfusion. These excessive ROS levels can lead to lipid peroxidation in cell membranes or trigger inflammatory responses, causing

cellular damage [54, 55]. Moreover, the surge in ROS amplifies oxidative stress, promotes apoptosis, and intensifies the severity of MIRI [56, 57]. Beyond contributing to oxidative stress, ROS also play a pivotal role in pyroptosis during MIRI. Shen et al. discovered that ROS could directly activate the NLRP3 inflammasome, thereby inducing pyroptosis [58]. Furthermore, ROS facilitates the activation and production of IL-18, which in turn promotes tissue inflammation and aggravates MIRI [59]. Pyroptosis itself can enhance ROS production through various mechanisms, including the disruption and impairment of mitochondrial function. Consequently, the interplay between pyroptosis and oxidative stress significantly contributes to the onset and progression of MIRI.

Pyroptosis and calcium overload

Calcium overload, often triggered by sodium pump dysfunction and an increase in ROS, is both a common cause and consequence of MIRI. This condition can disrupt the oxidative phosphorylation cycle and mitochondrial membrane potential, a well-acknowledged pathogenic mechanism in MIRI [60]. Researches have established a close link between calcium overload and pyroptosis. For instance, Wang et al. discovered in a rat MIRI model that calcium overload could initiate cardiomyocyte pyroptosis via the NLRP3/caspase-1 pathway. The application of ginsenoside, targeting the retinoblastoma susceptibility gene (Rb1) to mitigate calcium overload, was found to decrease myocardial pyroptosis and cellular damage [61]. Similarly, Zhou et al. demonstrated that inhibiting calcium overload could lessen pyroptosis in glioblastoma cells [62]. While the precise mechanism through which calcium overload directly influences pyroptosis remains unclear, there is an undeniable link between the two in the context of MIRI. Future research should delve into the reciprocal regulatory mechanisms between calcium overload and pyroptosis.

Pyroptosis, apoptosis, and programmed necrosis

The pathogenesis of MIRI is multifaceted, with apoptosis, pyroptosis, and programmed necrosis playing significant roles in its development [63]. Numerous studies have highlighted how these distinct forms of cell death may interact and impact one another, suggesting the presence of “crosstalk” between pyroptosis, apoptosis, and programmed necrosis, wherein caspase-8 plays a pivotal

role [64-67]. As a protease, caspase-8 primarily governs apoptosis but is also known to mediate programmed necrosis [68, 69]. This dual function raises questions about the multifaceted roles caspase-8 may play in cell death processes. Recent research has further elucidated caspase-8’s critical role in the non-classical pathway of pyroptosis. Upregulation of caspase-8 can suppress the expression of GSDMD by inhibiting the formation of ASC and pyroptosis [70]. These findings underscore the complex interplay between pyroptosis, apoptosis, and programmed necrosis, indicating that their interactions may significantly contribute to the progression of MIRI. Future research should delve into the potential crossover, superposition, or feedback mechanisms among these modes of cell death, aiming to better understand their collective impact on the onset and evolution of MIRI.

Drugs for targeted inhibition of cell death and amelioration of MIRI

NLRP3 inhibitors

MCC950 is a small molecule known for its selective inhibition of the NLRP3 inflammasome, effectively preventing the activation conformation of NLRP3 and, consequently, inhibiting both NLRP3 activation and pyroptosis [71, 72]. In a porcine model of MIRI, various doses of MCC950 significantly reduced both the area of myocardial infarction and the inflammatory response [73].

Colchicine, traditionally used in the clinical management of gout, has recently been identified as an effective NLRP3 inhibitor, showing promise in the prevention and treatment of MIRI [74, 75]. Colchicine targets tubulin which is crucial for the localization and assembly of the NLRP3 inflammasome, by irreversibly binding to it, thereby inhibiting microtubule formation and NLRP3 activation [76-79]. Studies in an AMI rat model demonstrated that nanoparticles loaded with colchicine markedly decreased NLRP3 and pyroptosis-related protein expression, reducing infarct size [80].

Metformin, a primary medication for type 2 diabetes, has also shown efficacy beyond diabetes, particularly in ischemia-reperfusion injuries affecting cardiovascular, liver, and kidney tissues [81-83]. Research by Zhang et al. revealed that metformin mitigates the inflammatory response in MIRI by activating AMP-activated protein kinase (AMPK) and suppressing NLRP3 inflammasome activation,

thereby offering cardioprotective benefits [84]. Additionally, metformin postconditioning in isolated rat heart perfusion and neonatal rat MIRI models inhibited NLRP3 inflammasome activation and reduced the expression of inflammatory markers such as tumor necrosis factor- α , IL-6, and IL-1 β , subsequently decreasing the area of myocardial infarction [81].

INF4E (ethyl 2-((2-chlorophenyl)hydroxyl) methyl) acrylate), another effective NLRP3 inflammasome inhibitor, has been shown to reduce the activity of caspase-1 and NLRP3-ATPase [85, 86]. Mastrocola et al. demonstrated that pretreatment with INF4E diminished myocardial infarct size, improved myocardial contractility, and downregulated the expression of the NLRP3 inflammasome and GSDMD via the reperfusion injury salvage kinase pathway, thereby alleviating MIRI [85].

These findings underscore the potential of targeting NLRP3 inhibition as a key strategy for preventing and treating MIRI. However, further research is necessary to develop more specific, safer, and efficacious NLRP3 inflammasome inhibitors, alongside large-scale clinical trials to verify their clinical safety and effectiveness.

GSDMD inhibitors

GSDMD serves as the executor and pivotal effector molecule in pyroptosis, playing a crucial role in the release of downstream inflammatory factors such as IL-18 and IL-1 β [87]. Consequently, compounds that target GSDMD have been developed, demonstrating potential therapeutic value across various disease models. Necrosulfonamide (NSA) is one such GSDMD inhibitor that binds directly to the C191 site of GSDMD, preventing its oligomerization and inhibiting its expression [88]. He et al. demonstrated that NSA treatment could suppress GSDMD expression and pyroptosis, thereby ameliorating cardiac dysfunction following ischemia/reperfusion injury in rats [89].

Another GSDMD inhibitor, dimethyl fumarate (DMF), acts by succinylating the C191 site of GSDMD, which also prevents its oligomerization [90]. Shi et al. found that DMF could diminish NLRP3 inflammasome activation and GSDMD expression through the regulation of protein kinase A signaling, reducing pyroptosis and the inflammatory response and offering relief in mouse models of autoimmune hepatitis [91]. While DMF's anti-inflammatory properties have been documented in various

conditions, including LPS-induced sepsis, familial Mediterranean fever, and autoimmune encephalitis, its effects on MIRI have yet to be explored [92].

Trimetazidine, a known inhibitor of free fatty acid oxidation, offers myocardial protection in patients with diabetes, angina, and those undergoing PCI [93, 94]. A recent study suggests that trimetazidine can also target GSDMD to modulate pyroptosis. It plays a significant role in regulating the expression of the TLR4/Myd88/NF- κ B pathway and the NLRP3 inflammasome [95].

Given GSDMD's central role in controlling the release of inflammatory factors during pyroptosis, targeting this molecule presents a promising approach to inhibit pyroptosis.

Caspase-1 inhibitors

Intriguingly, a clinical trial demonstrated that colchicine significantly dampens the activity of caspase-1 in monocytes and diminishes the secretion of IL-1 β by these cells, thereby delivering cardioprotective benefits [75]. VX-765, a highly selective inhibitor of caspase-1, has shown promise in a mouse model of MIRI [96, 97]. The concurrent use of VX-765 with antiplatelet medications markedly decreased myocardial infarct size and enhanced cardiac systolic function [98].

Further researches have revealed that VX-765 not only reduces the secretion of IL-1 β but also inhibits the release of lactate dehydrogenase, thereby mitigating MIRI [98, 99]. Carmo et al. discovered that VX-765 also shields isolated rat hearts via the reperfusion injury salvage kinase pathway [100, 101]. Additionally, α -1 antitrypsin (A1AT), another caspase-1 inhibitor, has been observed to downregulate caspase-1 expression upon intraperitoneal injection in a mouse MIRI model, significantly shrinking the area of myocardial infarction and enhancing ventricular remodeling [102-104].

These findings underscore that with a deeper understanding of the pyroptosis mechanism and the ongoing development of novel small-molecule drugs, a new generation of compounds targeting pyroptosis-specific mechanisms and MIRI is on the horizon. The advent of small-molecule compounds adept at specifically inhibiting pyroptosis holds substantial promise for the prevention and treatment of MIRI, heralding a significant advancement in cardiovascular disease management.

TCM compounds and monomers

Due to its holistic approach, encompassing multiple components, pathways, and targets, traditional Chinese medicine offers distinctive advantages in treating cardiovascular and cerebrovascular diseases, especially myocardial ischemia-reperfusion injury. Emodin, known for its anti-inflammatory and antioxidant properties, also inhibits GSDMD-mediated pyroptosis via the TLR4/MyD88/NF- κ B/NLRP3 inflammasome pathway, thereby reducing myocardial cell damage in MIRI [104, 105]. Researches have highlighted that naringin and β -asarone can diminish pyroptosis by suppressing the expression of ASC, caspase-1, NLRP3, and GSDMD, consequently reducing infarct size in rat models of MIRI [106, 107]. Similarly, the cinnamoyl ethyl acetate extract offers protection against MIRI damage in rats by inhibiting the activation of the NLRP3 inflammasome and pyroptosis [108]. Li et al. demonstrated that apigenin treatment lowers the expression of caspase-1, NLRP3, and GSDMD in cardiomyocytes subjected to hypoxia/reoxygenation (H/R), decreasing pyroptosis incidence and providing myocardial protection [109]. Sun et al. found that pre-treatment with gastrodin in cardiac microvascular endothelial cells can obstruct pyroptosis via the classical pathway, alleviate inflammation, and reduce myocardial cell damage [110].

These studies illustrate that Chinese medicines and their monomers possess significant potential in combating cell pyroptosis and MIRI due to their pharmacological actions. Nonetheless, future efforts should aim at a more thorough investigation and translational research on the modulation of pyroptosis and the protective effects of traditional Chinese medicine against MIRI, paving the way for their early clinical adoption.

Anesthetic

In recent years, the cardioprotective effects of anesthetics on cardiovascular and cerebrovascular diseases have garnered increasing attention from researchers globally. Both intravenous and inhalational anesthetics have been shown to offer myocardial protection [111, 112]. Sevoflurane, a widely used inhalational anesthetic in clinical settings, has been recognized for its cardioprotective properties in various studies [110].

Dharmalingam et al. reported that pre-

treatment with volatile anesthetics, including isoflurane and sevoflurane, can suppress ROS, thereby preventing oxidative stress during coronary artery bypass grafting [113]. Additional researches indicate that sevoflurane can also block the P2X7 ion channel on mitochondrial membranes, decrease the expression of NLRP3, ASC, caspase-1, and GSDMD, and reduce myocardial enzyme release, thereby playing a role in mitigating MIRI [114-116]. Deng et al. further revealed that sevoflurane can modulate autophagy via the AMPK/ULK1 pathway and inhibit NLRP3-mediated pyroptosis in cardiomyocytes, thus improving myocardial ischemia/reperfusion injury [117].

Dexmedetomidine, a highly selective α -2 adrenergic receptor agonist commonly utilized for perioperative sedation, has been found to protect against ischemia-reperfusion injury by enhancing myocardial function [118-120]. Zhong et al. and Wang et al. discovered that dexmedetomidine could mitigate MIRI in a rat model by downregulating miR-29b, activating FoxO3a, and reducing pyroptosis [121, 122]. While there is significant evidence to suggest that both inhaled and intravenous anesthetics have profound effects on the prevention and treatment of MIRI, there remains a scarcity of studies and mechanistic insights into how anesthetics improve MIRI by regulating pyroptosis [114, 123]. Further investigations into the specific targets and mechanisms by which anesthesia influences MIRI are crucial for advancing our understanding and therapeutic approaches.

Summary and outlook

MIRI is a multifaceted pathophysiological phenomenon influenced by numerous genes, molecules, cells, and tissues across different signaling pathways. This review delves into the role and mechanisms of pyroptosis in MIRI, summarizing the impact of various drug categories, including NLRP3 inhibitors, GSDMD inhibitors, caspase-1 inhibitors, anesthetics, and traditional Chinese medicines, on both MIRI and pyroptosis. Key pathogenic mechanisms of MIRI, such as calcium overload, disturbances in energy metabolism, oxidative stress, and autophagy, have been thoroughly investigated, highlighting their importance in modulating the onset and progression of MIRI. The contribution of pyroptosis to MIRI, especially the interactions between pyroptosis, MIRI mechanisms, and associated signaling pathways, remains a critical area for further research.

Most existing research on pyroptosis and MIRI concentrates on mitigating pyroptosis and improving MIRI outcomes by targeting the NLRP3 inflammasome, caspase family proteins, and key GSDMD targets. While the use of pyroptosis inhibitors in animal models of MIRI has shown promising results in reducing myocardial infarct size and enhancing cardiac function, the majority of these studies are confined to preclinical settings. The clinical application of drugs that inhibit pyroptosis-related proteins for MIRI treatment is limited, which restricts our comprehensive understanding of these inhibitors' effects in human clinical trials. Consequently, there is a pressing need for more clinical trials to corroborate the findings from basic research.

Based on current studies and available evidence, NLRP3 inhibitors, caspase-1 inhibitors, and traditional Chinese medicines emerge as potential therapeutic avenues for MIRI treatment. However, given the intricate nature of pyroptosis and MIRI, a singular focus on targeting either pyroptosis or MIRI may not suffice for effective prevention and treatment strategies. Instead, a multi-targeted and multi-mechanism approach to combination therapy may offer a more effective treatment paradigm for MIRI.

Author Contributions: Ziyue Li helped draft the manuscript. Bailong Hu and Xiaohua Zou revised the manuscript. All authors have read and approved the final draft.

References

- [1] Anderson JL, Morrow DA. Acute Myocardial Infarction. *N Engl J Med* 2017;376(21):2053-2064.
- [2] Johansson S, Rosengren A, Young K, et al. Mortality and morbidity trends after the first year in survivors of acute myocardial infarction: a systematic review. *BMC Cardiovasc Disord* 2017;17(1):53.
- [3] Kapur NK, Thayer KL, Zweck E. Cardiogenic Shock in the Setting of Acute Myocardial Infarction. *Methodist Debakey Cardiovasc J* 2020;16(1):16-21.
- [4] Devkota S, Dhungana RR, Pandey AR, et al. Risk Factors of Coronary Artery Disease: A Hospital-Based Study. *J Nepal Health Res Counc* 2022;20(2):487-493.
- [5] Bhatt DL. Percutaneous Coronary Intervention in 2018. *Jama* 2018;319(20):2127-2128.
- [6] Gaudino M, Benedetto U, Fremes S, et al. Association of Radial Artery Graft vs Saphenous Vein Graft With Long-term Cardiovascular Outcomes Among Patients Undergoing Coronary Artery Bypass Grafting: A Systematic Review and Meta-analysis. *Jama* 2020;324(2):179-187.
- [7] Weidenmann V, Robinson NB, Rong LQ, et al. Diagnostic dilemma of perioperative myocardial infarction after coronary artery bypass grafting: A review. *Int J Surg* 2020;79:76-83.
- [8] Hausenloy DJ, Yellon DM. Myocardial ischemia-reperfusion injury: a neglected therapeutic target. *J Clin Invest* 2013;123(1):92-100.
- [9] Tanzilli G, Truscelli G, Arrivi A, et al. Glutathione infusion before primary percutaneous coronary intervention: a randomised controlled pilot study. *BMJ Open* 2019;9(8):e025884.
- [10] Zhao L, Xing C, Yang P, et al. Echocardiographic observation on myocardial ischemia reperfusion injury after percutaneous coronary intervention. *Chin J Med Imaging Technol* 2020;36(1):5.
- [11] Luo Y, Lyu L, Li G, et al. Analysis on correlative factors for occurrence of myocardial ischemia-reperfusion injury during primary percutaneous coronary intervention for acute myocardial infarction. *Chin J Cardiol* 2005;33(8):4.
- [12] Wu W, Chen X, Hu Q, et al. Improvement of Myocardial Cell Injury by miR-199a-3p/mTOR Axis through Regulating Cell Apoptosis and Autophagy. *J Immunol Res* 2022;2022:1642301.
- [13] Yang CF. Clinical manifestations and basic mechanisms of myocardial ischemia/reperfusion injury. *Ci Ji Yi Xue Za Zhi* 2018;30(4):209-215.
- [14] Bell RM, Bøtker HE, Carr RD, et al. 9th Hatter Biannual Meeting: position document on ischaemia/reperfusion injury, conditioning and the ten commandments of cardioprotection. *Basic Res Cardiol* 2016;111(4):41.
- [15] Davidson SM, Ferdinandy P, Andreadou I, et al. Multitarget Strategies to Reduce Myocardial Ischemia/Reperfusion Injury: JACC Review Topic of the Week. *J Am Coll Cardiol* 2019;73(1):89-99.
- [16] Kovacs SB, Miao EA. Gasdermins: Effectors of Pyroptosis. *Trends Cell Biol* 2017;27(9):673-684.
- [17] Zhaolin Z, Guohua L, Shiyuan W, et al. Role of pyroptosis in cardiovascular disease. *Cell Prolif* 2019;52(2):e12563.
- [18] Friedlander AM. Macrophages are sensitive to anthrax lethal toxin through an acid-dependent process. *J Biol Chem* 1986;261(16):7123-7126.
- [19] Zychlinsky A, Prevost MC, Sansonetti PJ.

- Shigella flexneri* induces apoptosis in infected macrophages. *Nature* 1992;358(6382):167-169.
- [20] D'Souza CA, Heitman J. Dismantling the *Cryptococcus* coat. *Trends Microbiol* 2001;9(3):112-113.
- [21] Martinon F, Burns K, Tschopp J. The inflammasome: a molecular platform triggering activation of inflammatory caspases and processing of proIL-beta. *Mol Cell* 2002;10(2):417-426.
- [22] Sun S, Gong D, Liu R, et al. Puerarin Inhibits NLRP3-Caspase-1-GSDMD-Mediated Pyroptosis via P2X7 Receptor in Cardiomyocytes and Macrophages. *Int J Mol Sci* 2023;24(17):13169.
- [23] Long J, Sun Y, Liu S, et al. Targeting pyroptosis as a preventive and therapeutic approach for stroke. *Cell Death Discov* 2023;9(1):155.
- [24] Galluzzi L, Vitale I, Aaronson SA, et al. Molecular mechanisms of cell death: recommendations of the Nomenclature Committee on Cell Death 2018. *Cell Death Differ* 2018;25(3):486-541.
- [25] Cookson BT, Brennan MA. Pro-inflammatory programmed cell death. *Trends Microbiol* 2001;9(3):113-114.
- [26] Yu P, Zhang X, Liu N, et al. Pyroptosis: mechanisms and diseases. *Signal Transduct Target Ther* 2021;6(1):128.
- [27] Strowig T, Henao-Mejia J, Elinav E, et al. Inflammasomes in health and disease. *Nature* 2012;481(7381):278-286.
- [28] Liston A, Masters SL. Homeostasis-altering molecular processes as mechanisms of inflammasome activation. *Nat Rev Immunol* 2017;17(3):208-214.
- [29] He Y, Hara H, Núñez G. Mechanism and Regulation of NLRP3 Inflammasome Activation. *Trends Biochem Sci* 2016;41(12):1012-1021.
- [30] Kelley N, Jeltema D, Duan Y, et al. The NLRP3 Inflammasome: An Overview of Mechanisms of Activation and Regulation. *Int J Mol Sci* 2019;20(13):3328.
- [31] Huang Y, Xu W, Zhou R. NLRP3 inflammasome activation and cell death. *Cell Mol Immunol* 2021;18(9):2114-2127.
- [32] Zhang M, Xin W, Yu Y, et al. Programmed death-ligand 1 triggers PSMCs pyroptosis and pulmonary vascular fibrosis in pulmonary hypertension. *J Mol Cell Cardiol* 2020;138:23-33.
- [33] Ding J, Wang K, Liu W, et al. Pore-forming activity and structural autoinhibition of the gasdermin family. *Nature* 2016;535(7610):111-116.
- [34] Chen X, He WT, Hu L, et al. Pyroptosis is driven by non-selective gasdermin-D pore and its morphology is different from MLKL channel-mediated necroptosis. *Cell Res* 2016;26(9):1007-20.
- [35] Sborgi L, Rühl S, Mulvihill E, et al. GSDMD membrane pore formation constitutes the mechanism of pyroptotic cell death. *Embo j* 2016;35(16):1766-1778.
- [36] Bergsbaken T, Fink SL, Cookson BT. Pyroptosis: host cell death and inflammation. *Nat Rev Microbiol* 2009;7(2):99-109.
- [37] Shi J, Zhao Y, Wang K, et al. Cleavage of GSDMD by inflammatory caspases determines pyroptotic cell death. *Nature* 2015;526(7575):660-665.
- [38] He WT, Wan H, Hu L, et al. Gasdermin D is an executor of pyroptosis and required for interleukin-1 β secretion. *Cell Res* 2015;25(12):1285-1298.
- [39] Shi J, Zhao Y, Wang Y, et al. Inflammatory caspases are innate immune receptors for intracellular LPS. *Nature* 2014;514(7521):187-192.
- [40] Aglietti RA, Estevez A, Gupta A, et al. GsdmD p30 elicited by caspase-11 during pyroptosis forms pores in membranes. *Proc Natl Acad Sci U S A* 2016;113(28):7858-7863.
- [41] Baker PJ, Boucher D, Bierschenk D, et al. NLRP3 inflammasome activation downstream of cytoplasmic LPS recognition by both caspase-4 and caspase-5. *Eur J Immunol* 2015;45(10):2918-2926.
- [42] Rühl S, Broz P. Caspase-11 activates a canonical NLRP3 inflammasome by promoting K(+) efflux. *Eur J Immunol* 2015;45(10):2927-2936.
- [43] Shi J, Gao W, Shao F. Pyroptosis: Gasdermin-Mediated Programmed Necrotic Cell Death. *Trends Biochem Sci* 2017;42(4):245-254.
- [44] Wang Y, Gao W, Shi X, et al. Chemotherapy drugs induce pyroptosis through caspase-3 cleavage of a gasdermin. *Nature* 2017;547(7661):99-103.
- [45] Liu Y, Fang Y, Chen X, et al. Gasdermin E-mediated target cell pyroptosis by CAR T cells triggers cytokine release syndrome. *Sci Immunol* 2020; 17;5(43).
- [46] Hou J, Zhao R, Xia W, et al. PD-L1-mediated gasdermin C expression switches apoptosis to pyroptosis in cancer cells and facilitates tumour necrosis. *Nat Cell Biol* 2020;22(10):1264-1275.
- [47] Xu XN, Jiang Y, Yan LY, et al. Aesculin suppresses the NLRP3 inflammasome-mediated pyroptosis via the Akt/GSK3 β /NF- κ B pathway to mitigate myocardial ischemia/reperfusion injury. *Phytomedicine* 2021;92:153687.
- [48] Sandanger Ø, Ranheim T, Vinge LE, et al. The NLRP3 inflammasome is up-regulated in cardiac fibroblasts and mediates myocardial ischaemia-reperfusion injury. *Cardiovasc Res*

- 2013;99(1):164-74.
- [49] Zhang B, Liu G, Huang B, et al. KDM3A Attenuates Myocardial Ischemic and Reperfusion Injury by Ameliorating Cardiac Microvascular Endothelial Cell Pyroptosis. *Oxid Med Cell Longev* 2022;2022:4622520.
- [50] Dai Y, Wang S, Chang S, et al. M2 macrophage-derived exosomes carry microRNA-148a to alleviate myocardial ischemia/reperfusion injury via inhibiting TXNIP and the TLR4/NF- κ B/NLRP3 inflammasome signaling pathway. *J Mol Cell Cardiol* 2020;142:65-79.
- [51] Wang Y, Qiu Z, Yuan J, et al. Hypoxia-reoxygenation induces macrophage polarization and causes the release of exosomal miR-29a to mediate cardiomyocyte pyroptosis. *In Vitro Cell Dev Biol Anim* 2021;57(1):30-41.
- [52] Wang R, Liu F, Huang P, et al. Ozone preconditioning protects rabbit heart against global ischemia-reperfusion injury in vitro by up-regulating HIF-1 α . *Biomed Pharmacother* 2022;150:113033.
- [53] Tian Y, Li H, Liu P, et al. Captopril Pretreatment Produces an Additive Cardioprotection to Isoflurane Preconditioning in Attenuating Myocardial Ischemia Reperfusion Injury in Rabbits and in Humans. *Mediators Inflamm* 2015;2015:819232.
- [54] Zhan KY, Yu PL, Liu CH, et al. Detrimental or beneficial: the role of TRPM2 in ischemia/reperfusion injury. *Acta Pharmacol Sin* 2016;37(1):4-12.
- [55] Qiu Y, Shi YN, Zhu N, et al. A Lipid Perspective on Regulated Pyroptosis. *Int J Biol Sci* 2023;19(8):2333-2348.
- [56] Kurian GA, Rajagopal R, Vedantham S, et al. The Role of Oxidative Stress in Myocardial Ischemia and Reperfusion Injury and Remodeling: Revisited. *Oxid Med Cell Longev* 2016;2016:1656450.
- [57] González-Montero J, Brito R, Gajardo AI, et al. Myocardial reperfusion injury and oxidative stress: Therapeutic opportunities. *World J Cardiol* 2018;10(9):74-86.
- [58] Shen S, He F, Cheng C, et al. Uric acid aggravates myocardial ischemia-reperfusion injury via ROS/NLRP3 pyroptosis pathway. *Biomed Pharmacother* 2021;133:110990.
- [59] Li H, Yang DH, Zhang Y, et al. Geniposide suppresses NLRP3 inflammasome-mediated pyroptosis via the AMPK signaling pathway to mitigate myocardial ischemia/reperfusion injury. *Chin Med* 2022;17(1):73.
- [60] Qian W, Xiong X, Fang Z, et al. Protective effect of tetramethylpyrazine on myocardial ischemia-reperfusion injury. *Evid Based Complement Alternat Med* 2014;2014:107501.
- [61] Wang M, Wang R, Sun H, et al. Ginsenoside Rb1 ameliorates cardiotoxicity triggered by aconitine via inhibiting calcium overload and pyroptosis. *Phytomedicine* 2021;83:153468.
- [62] Zhou B, Lin Y, Chen S, et al. Activation of Ca(2+)/Calmodulin-Dependent Protein Kinase II (CaMKII) with Lidocaine Provokes Pyroptosis of Glioblastoma Cells. *Bull Exp Biol Med* 2021;171(3):297-304.
- [63] Bertheloot D, Latz E, Franklin BS. Necroptosis, pyroptosis and apoptosis: an intricate game of cell death. *Cell Mol Immunol* 2021;18(5):1106-1121.
- [64] Fritsch M, Günther SD, Schwarzer R, et al. Caspase-8 is the molecular switch for apoptosis, necroptosis and pyroptosis. *Nature* 2019;575(7784):683-687.
- [65] Chen KW, Demarco B, Heilig R, et al. Extrinsic and intrinsic apoptosis activate pannexin-1 to drive NLRP3 inflammasome assembly. *Embo J* 2019;38(10):e101638.
- [66] Orning P, Weng D, Starheim K, et al. Pathogen blockade of TAK1 triggers caspase-8-dependent cleavage of gasdermin D and cell death. *Science* 2018;362(6418):1064-1069.
- [67] Sarhan J, Liu BC, Muendlein HI, et al. Caspase-8 induces cleavage of gasdermin D to elicit pyroptosis during Yersinia infection. *Proc Natl Acad Sci U S A* 2018;115(46):E10888-e10897.
- [68] Feng S, Yang Y, Mei Y, et al. Cleavage of RIP3 inactivates its caspase-independent apoptosis pathway by removal of kinase domain. *Cell Signal* 2007;19(10):2056-67.
- [69] Newton K, Wickliffe KE, Dugger DL, et al. Cleavage of RIPK1 by caspase-8 is crucial for limiting apoptosis and necroptosis. *Nature* 2019;574(7778):428-431.
- [70] Vince JE, De Nardo D, Gao W, et al. The Mitochondrial Apoptotic Effectors BAX/BAK Activate Caspase-3 and -7 to Trigger NLRP3 Inflammasome and Caspase-8 Driven IL-1 β Activation. *Cell Rep* 2018;25(9):2339-2353. e4.
- [71] Grant AJ, Yang N, Moore MJ, et al. Selective NLRP3 Inflammasome Inhibitor MCC950 Suppresses Inflammation and Facilitates Healing in Vascular Materials. *Adv Sci (Weinh)* 2023;10(20):e2300521.
- [72] Wu D, Chen Y, Sun Y, et al. Target of MCC950 in Inhibition of NLRP3 Inflammasome Activation: a Literature Review. *Inflammation* 2020;43(1):17-23.
- [73] van Hout GP, Bosch L, Ellenbroek GH, et al. The selective NLRP3-inflammasome inhibitor MCC950 reduces infarct size and preserves cardiac function in a pig model of myocardial infarction. *Eur Heart J* 2017;38(11):828-836.
- [74] Bonaventura A, Vecchié A, Dagna L, et al. Colchicine for COVID-19: targeting NLRP3 inflammasome to blunt hyperinflammation. *Inflamm Res* 2022;71(3):293-307.

- [75] Robertson S, Martínez GJ, Payet CA, et al. Colchicine therapy in acute coronary syndrome patients acts on caspase-1 to suppress NLRP3 inflammasome monocyte activation. *Clin Sci (Lond)* 2016;130(14):1237-46.
- [76] Li CG, Zeng QZ, Chen MY, et al. Evodiamine Augments NLRP3 Inflammasome Activation and Anti-bacterial Responses Through Inducing α -Tubulin Acetylation. *Front Pharmacol* 2019;10:290.
- [77] Nolasco S, Bellido J, Serna M, et al. Colchicine Blocks Tubulin Heterodimer Recycling by Tubulin Cofactors TBCA, TBCB, and TBCE. *Front Cell Dev Biol* 2021;9:656273.
- [78] Zhong B, Sun S, Tan KS, et al. Hypoxia-inducible factor 1 α activates the NLRP3 inflammasome to regulate epithelial differentiation in chronic rhinosinusitis. *J Allergy Clin Immunol* 2023;152(6):1444-1459.e14.
- [79] Demidowich AP, Davis AI, Dedhia N, et al. Colchicine to decrease NLRP3-activated inflammation and improve obesity-related metabolic dysregulation. *Med Hypotheses* 2016;92:67-73.
- [80] Wang L, Peng Y, Song L, et al. Colchicine-Containing Nanoparticles Attenuates Acute Myocardial Infarction Injury by Inhibiting Inflammation. *Cardiovasc Drugs Ther* 2022;36(6):1075-1089.
- [81] Fei Q, Ma H, Zou J, et al. Metformin protects against ischaemic myocardial injury by alleviating autophagy-ROS-NLRP3-mediated inflammatory response in macrophages. *J Mol Cell Cardiol* 2020;145:1-13.
- [82] Triggler CR, Mohammed I, Bshesh K, et al. Metformin: Is it a drug for all reasons and diseases? *Metabolism* 2022;133:155223.
- [83] Foretz M, Guigas B, Bertrand L, et al. Metformin: from mechanisms of action to therapies. *Cell Metab* 2014;20(6):953-66.
- [84] Zhang J, Huang L, Shi X, et al. Metformin protects against myocardial ischemia-reperfusion injury and cell pyroptosis via AMPK/NLRP3 inflammasome pathway. *Aging (Albany NY)* 2020;12(23):24270-24287.
- [85] Mastrocola R, Penna C, Tullio F, et al. Pharmacological Inhibition of NLRP3 Inflammasome Attenuates Myocardial Ischemia/Reperfusion Injury by Activation of RISK and Mitochondrial Pathways. *Oxid Med Cell Longev* 2016;2016:5271251.
- [86] Shi Y, Lv Q, Zheng M, et al. NLRP3 inflammasome inhibitor INF39 attenuated NLRP3 assembly in macrophages. *Int Immunopharmacol* 2021;92:107358.
- [87] Swanson KV, Deng M, Ting JP. The NLRP3 inflammasome: molecular activation and regulation to therapeutics. *Nat Rev Immunol* 2019;19(8):477-489.
- [88] Rathkey JK, Zhao J, Liu Z, et al. Chemical disruption of the pyroptotic pore-forming protein gasdermin D inhibits inflammatory cell death and sepsis. *Sci Immunol* 2018;3(26):eaat2738.
- [89] He F, Zheng G, Hu J, et al. Necrosulfonamide improves post-resuscitation myocardial dysfunction via inhibiting pyroptosis and necroptosis in a rat model of cardiac arrest. *Eur J Pharmacol* 2022;926:175037.
- [90] Li N, Chen J, Geng C, et al. Myoglobin promotes macrophage polarization to M1 type and pyroptosis via the RIG-I/Caspase1/GSDMD signaling pathway in CS-AKI. *Cell Death Discov* 2022;8(1):90.
- [91] Shi FL, Ni ST, Luo SQ, et al. Dimethyl fumarate ameliorates autoimmune hepatitis in mice by blocking NLRP3 inflammasome activation. *Int Immunopharmacol* 2022;108:108867.
- [92] Coll RC, Schroder K, Pelegrin P. NLRP3 and pyroptosis blockers for treating inflammatory diseases. *Trends Pharmacol Sci* 2022;43(8):653-668.
- [93] Marzilli M, Vinereanu D, Lopaschuk G, et al. Trimetazidine in cardiovascular medicine. *Int J Cardiol* 2019;293:39-44.
- [94] Ferrari R, Ford I, Fox K, et al. Efficacy and safety of trimetazidine after percutaneous coronary intervention (ATPCI): a randomised, double-blind, placebo-controlled trial. *Lancet* 2020;396(10254):830-838.
- [95] Chen X, Lin S, Dai S, et al. Trimetazidine affects pyroptosis by targeting GSDMD in myocardial ischemia/reperfusion injury. *Inflamm Res* 2022;71(2):227-241.
- [96] McKenzie BA, Mamik MK, Saito LB, et al. Caspase-1 inhibition prevents glial inflammasome activation and pyroptosis in models of multiple sclerosis. *Proc Natl Acad Sci U S A* 2018;115(26):E6065-e6074.
- [97] Yang XM, Downey JM, Cohen MV, et al. The Highly Selective Caspase-1 Inhibitor VX-765 Provides Additive Protection Against Myocardial Infarction in Rat Hearts When Combined With a Platelet Inhibitor. *J Cardiovasc Pharmacol Ther* 2017;22(6):574-578.
- [98] Audia JP, Yang XM, Crockett ES, et al. Caspase-1 inhibition by VX-765 administered at reperfusion in P2Y(12) receptor antagonist-treated rats provides long-term reduction in myocardial infarct size and preservation of ventricular function. *Basic Res Cardiol* 2018;113(5):32.
- [99] Rout A, Tantry US, Novakovic M, et al. Targeted pharmacotherapy for ischemia reperfusion injury in acute myocardial infarction. *Expert Opin Pharmacother* 2020;21(15):1851-1865.
- [100] Do Carmo H, Arjun S, Petrucci O, et al. The Caspase 1 Inhibitor VX-765 Protects the

- Isolated Rat Heart via the RISK Pathway. *Cardiovasc Drugs Ther* 2018;32(2):165-168.
- [101] Rossello X, Yellon DM. The RISK pathway and beyond. *Basic Res Cardiol* 2018;113(1):2.
- [102] Jiang D, Berman R, Wu Q, et al. The Anti-inflammatory Effect of Alpha-1 Antitrypsin in Rhinovirus-infected Human Airway Epithelial Cells. *J Clin Cell Immunol* 2016;7(6):475.
- [103] Mauro AG, Mezzaroma E, Marchetti C, et al. A Preclinical Translational Study of the Cardioprotective Effects of Plasma-Derived Alpha-1 Anti-trypsin in Acute Myocardial Infarction. *J Cardiovasc Pharmacol* 2017;69(5):273-278.
- [104] Toldo S, Seropian IM, Mezzaroma E, et al. Alpha-1 antitrypsin inhibits caspase-1 and protects from acute myocardial ischemia-reperfusion injury. *J Mol Cell Cardiol* 2011;51(2):244-251.
- [105] Semwal RB, Semwal DK, Combrinck S, et al. Emodin - A natural anthraquinone derivative with diverse pharmacological activities. *Phytochemistry* 2021;190:112854.
- [106] Wang T, Zhang J, Zhang Z, et al. Naringin inhibits pyroptosis induced by myocardial ischemia/reperfusion injury in rats. *Chin J Pathophysiol* 2021;37(6):1019-1026.
- [107] Xiao B, Huang X, Wang Q, et al. Beta-Asarone Alleviates Myocardial Ischemia-Reperfusion Injury by Inhibiting Inflammatory Response and NLRP3 Inflammasome Mediated Pyroptosis. *Biol Pharm Bull* 2020;43(7):1046-1051.
- [108] Peng L, Lei Z, Rao Z, et al. Cardioprotective activity of ethyl acetate extract of *Cinnamomi Ramulus* against myocardial ischemia/reperfusion injury in rats via inhibiting NLRP3 inflammasome activation and pyroptosis. *Phytomedicine* 2021;93:153798.
- [109] Li W, Chen L, Xiao Y. Apigenin protects against ischemia-/hypoxia-induced myocardial injury by mediating pyroptosis and apoptosis. *In Vitro Cell Dev Biol Anim* 2020;56(4):307-312.
- [110] Sun W, Lu H, Lyu L, et al. Gastrodin ameliorates microvascular reperfusion injury-induced pyroptosis by regulating the NLRP3/caspase-1 pathway. *J Physiol Biochem* 2019;75(4):531-547.
- [111] Lemoine S, Tritapepe L, Hanouz JL, et al. The mechanisms of cardio-protective effects of desflurane and sevoflurane at the time of reperfusion: anaesthetic post-conditioning potentially translatable to humans? *Br J Anaesth* 2016;116(4):456-475.
- [112] Landoni G, Lomivorotov VV, Nigro Neto C, et al. Volatile Anesthetics versus Total Intravenous Anesthesia for Cardiac Surgery. *N Engl J Med* 2019;380(13):1214-1225.
- [113] Dharmalingam SK, Amirtharaj GJ, Ramachandran A, et al. Volatile anesthetic preconditioning modulates oxidative stress and nitric oxide in patients undergoing coronary artery bypass grafting. *Ann Card Anaesth* 2021;24(3):319-326.
- [114] Wu J, Cai W, Du R, et al. Sevoflurane Alleviates Myocardial Ischemia Reperfusion Injury by Inhibiting P2X7-NLRP3 Mediated Pyroptosis. *Front Mol Biosci* 2021;8:768594.
- [115] Shokoples BG, Paradis P, Schiffrin EL. P2X7 Receptors: An Untapped Target for the Management of Cardiovascular Disease. *Arterioscler Thromb Vasc Biol* 2021;41(1):186-199.
- [116] Zhou J, Zhou Z, Liu X, et al. P2X7 Receptor-Mediated Inflammation in Cardiovascular Disease. *Front Pharmacol* 2021;12:654425.
- [117] Deng L, Jiang L, Wei N, et al. Anesthetic sevoflurane simultaneously regulates autophagic flux and pyroptotic cell death-associated cellular inflammation in the hypoxic/re-oxygenated cardiomyocytes: Identification of sevoflurane as putative drug for the treatment of myocardial ischemia-reperfusion injury. *Eur J Pharmacol* 2022;936:175363.
- [118] Weerink MAS, Struys M, Hannivoort LN, et al. Clinical Pharmacokinetics and Pharmacodynamics of Dexmedetomidine. *Clin Pharmacokinet* 2017;56(8):893-913.
- [119] Yu P, Zhang J, Ding Y, et al. Dexmedetomidine post-conditioning alleviates myocardial ischemia-reperfusion injury in rats by ferroptosis inhibition via SLC7A11/GPX4 axis activation. *Hum Cell* 2022;35(3):836-848.
- [120] He H, Liu P, Li P. Dexmedetomidine Ameliorates Cardiac Ischemia/Reperfusion Injury by Enhancing Autophagy Through Activation of the AMPK/SIRT3 Pathway. *Drug Des Devel Ther* 2023;17:3205-3218.
- [121] Zhong Y, Li YP, Yin YQ, et al. Dexmedetomidine inhibits pyroptosis by down-regulating miR-29b in myocardial ischemia reperfusion injury in rats. *Int Immunopharmacol* 2020;86:106768.
- [122] Wang Z, Yao M, Jiang L, et al. Dexmedetomidine attenuates myocardial ischemia/reperfusion-induced ferroptosis via AMPK/GSK-3 β /Nrf2 axis. *Biomed Pharmacother* 2022;154:113572.
- [123] Zhang WY, Zhang QL, Xu MJ. Effects of propofol on myocardial ischemia reperfusion injury through inhibiting the JAK/STAT pathway. *Eur Rev Med Pharmacol Sci* 2019;23(14):6339-6345.